






Chemicompetent DH5 α Cell Transformation










Introduction

This protocol yields transformed DH5 α chemicompetent cells.

Reagents

-  BL21 electrocompetent cells
-  Plasmid DNA
-  SOB liquid media
-  LB/Kanamycin plates
-  H₂O

Equipment

-  Electroporator
-  Electroporation cuvette
-  Eppendorf tubes
-  250 mL beaker
-  Hot plate
-  Thermometer
-  Ice in a bucket
-  Pipette and tips
-  Incubation cabinet

Procedure

1. Thaw a tube of 50 μ L chemicompetent cells on ice.
2. Add 1-5 μ L of plasmid DNA to the thawed cells and mix them gently by pipetting the mix up and down. Place them on ice for 30 minutes. Incubate LB/Kanamycin plates at 37°C to warm them.
3. Heat shock cells by placing them into a 42°C bath for 30 seconds. Place on ice for 5 minutes.
4. Transfer the sample to an Eppendorf tube and add 950 μ L of SOB media. Incubate the media and sample at 37°C for 60 minutes. Shake vigorously at 250 rpm.
5. Mix cells without vortexing and spread 50-100 μ L of cells onto pre-warmed LB/Kanamycin plates. Incubate at 37°C overnight.